The Impact of a Low Glycemic Index Diet on Inflammatory Markers and Serum Adiponectin Concentration in Adolescent Overweight and Obese Girls: A Randomized Clinical Trial

Authors

M. H. Rouhani¹, R. Kelishadi², M. Hashemipour^{3, 4}, A. Esmaillzadeh^{5, 7}, P. J. Surkan⁶, A. Keshavarz⁸, I. Azadbakht^{5, 7}

Affiliations

Affiliation addresses are listed at the end of the article

Key words

- low glycemic index
- insulin resistance
- inflammation
- female

received 19.05.2015 accepted 05.01.2016

Bibliography

DOI http://dx.doi.org/ 10.1055/s-0042-100467 Horm Metab Res 2016; 48: 251–256 © Georg Thieme Verlag KG Stuttgart · New York

Correspondence

ISSN 0018-5043

L. Azadbakht, PhD

Department of Community Nutrition School of Nutrition and Food Science Isfahan University of Medical Sciences PO Box 81745 Isfahan Iran

Tel.: +98/311/7922 719 Fax: +98/311/6682 509 azadbakht@hlth.mui.ac.ir

Abstract

1

Although the effects of dietary glycemic index (GI) on insulin resistance are well documented in adults, the complex interaction among glucose intolerance, inflammatory markers, and adipokine concentration has not been well studied, especially among adolescents. We investigated the effect of a low glycemic index (LGI) diet on insulin concentration, fasting blood sugar (FBS), inflammatory markers, and serum adiponectin concentration among healthy obese/overweight adolescent females. In this parallel randomized clinical trial, 2 different diets, an LGI diet and a healthy nutritional recommendation diet (HNRD) with similar macronutrient composition were prescribed to 50 obese and overweight adolescent girls with the same pubertal status. Biochemical markers FBS, serum insulin concentration, high sensitivity C-reactive protein (hs-CRP), interleukin 6 (IL-6), and adiponectin were measured

before and after a 10 week intervention. Using an intention-to-treat analysis, data from 50 subjects were analyzed. According to a dietary assessment, GI in the LGI group was 43.22 ±0.54. While the mean for FBS, serum insulin concentration, the homeostasis model assessment (HOMA), the quantitative insulin sensitivity check index (QUICKI), and adiponectin concentration did not differ significantly within each group, the average hs-CRP and IL-6 decreased significantly in the LGI diet group after the 10 week intervention (p=0.009 and p=0.001; respectively). Comparing percent changes, we found a marginally significant decrease in hs-CRP in the LGI group compared with the HNRD group after adjusting for confounders. Compliance with an LGI diet may have favorable effect on inflammation among overweight and obese adolescent girls.

Supporting Information for this article is available online at http://www.thieme-connect.de/products

Abbreviations

 \blacksquare

FBS Fasting blood sugar GI Glycemic index HGI High glycemic index

HNRD Healthy nutritional recommendation diet

hs-CRP High sensitivity C-reactive protein

IL-6 Interleukin 6LGI Low glycemic index

QUICKI Quantitative insulin sensitivity check index

Introduction

 $\overline{\mathbf{w}}$

Insulin resistance, a major cause of diabetes mellitus and metabolic syndrome [1,2], is usually associated with obesity [3]. It is also closely associated with serum adiponectin concentration [4–6] and inflammatory cytokines such as high

sensitivity C-reactive protein (hs-CRP) [1]. This relationship between blood glucose, insulin resistance, and inflammatory cytokines, however, does not appear to depend on obesity status [7.8]. Previous studies show that serum adiponectin concentration has an inverse correlation with obesity [9] and concentration of inflammatory markers [e.g., interleukin 6 (IL-6) and hs-CRP] [4]. Obesity usually results in chronic inflammation [10]. The association between low-grade inflammation and obesity has also been observed among adolescents living in developing countries [11]. Therefore, it is important to identify effective nutritional interventions to improve insulin resistance and inflammation in overweight and obese adolescents in such populations.

Although it is well established that adiponectin concentration, insulin resistance, and inflammatory markers are influenced by environmental conditions, particularly diet [12-14], the role of carbohydrate quality has not been emphasized. The potential of different foods to increase postprandial blood sugar has been defined as the glycemic index (GI). Previous studies reported that dietary GI may affekt insulin resistance-related factors such as, insulin concentration, fasting blood sugar (FBS) [15], and inflammatory markers [16,17] in adults. An inverse association between GI and adiponectin has been reported among the elderly [18]. The effekt of dietary GI on insulin concentration and FBS has previously been examined among adolescents [19-22]. Although some of these studies measured changes in inflammatory markers [23,24], variations in adiponectin in adolescents have not been assessed following a dietary GI intervention. Few studies have examined the effekt of GI on insulin resistance related factors simultaneously, such as inflammatory markers and adiponectin among adolescents. Furthermore, prior studies have been conducted with both sexes combined and, therefore, physiological sex differences may have confounding role. Moreover, the prevalence of obesity among females is quite concerning [25]. Limited evidence has been reported regarding inflammation and adiponectin in adolescents. Therefore, the aim of the current study was to examine the effekt of a low glycemic index (LGI) diet on insulin concentration, FBS, inflammatory markers and adiponectin among healthy overweight and obese adolescent girls.

Subjects and Methods

.

Subjects

Eligible participants for this parallel randomized clinical trial were identified using recruitment flyers and Isfahanian student health booklets. An introductory meeting was held in which a detailed explanation was given to the volunteers' parents, and informed written consent was obtained. The study was conducted in Isfahan, Iran in 2011. Adolescents were eligible to participate if they were female, between 12 and 18 years old, overweight or obese, menstruating and not on medication. Low compliance with recommendations and use of medications that interact with inflammatory responses or insulin function were exclusion criteria. Based on WHO body mass index (BMI) definitions [26], overweight and obesity were defined as between 85th and 95th percentile and greater than the 95th percentile, respectively. Information on age, menstrual status and medication use were collected orally. Fifty adolescent participants were recruited for the study. The required sample size was calculated by $N = 2[(Z_{1-\alpha/2} + Z_{1-\beta})^2 \times S^2]/d^2$ [27] where $\alpha = 0.05$ (Type I error) and β = 0.20 (Type II error). The main outcome variable was hs-CRP. Variance of hs-CRP was 0.9 [23] and the difference in its mean was 1. The formula estimated that 13 subjects were required for each group. This study was approved by the Research Council and Ethical Committee of the School of Nutrition and Food Science, Isfahan University of Medical Sciences, Isfahan, Iran and the Food Security Research Center, Isfahan University of Medical Sciences, Isfahan, Iran. This randomized clinical trial was registered at IRCT.ir (IRCT201109272839N4).

Study procedure

Participants were randomly assigned to an LGI or healthy nutritional recommendation diet (HNRD) using a random number table. Study duration was 10 weeks. Although the biochemical

laboratory staffs were blinded, we could not blind the participants to the intervention type because it was a dietary intervention. In this study, there were 3 important variables (age, sex, and pubertal status) that were matched by restricting our inclusion of participants to: subjects who were female, menstruating, and between 12–18 years old. Clinical visits were scheduled each 2.5 weeks. Therefore, 4 sessions were conducted before the end of the study.

Dietary intervention

According to the US Institutes of Medicine formula [28], total energy expenditure was estimated for each participant. A moderate caloric restriction (200 kcal) was given only to adolescents with BMI > 95th percentile. Recommended macronutrient distribution was similar between the 2 groups (53–56% carbohydrate, 16-18% proteins and 27-30% fat). LGI and high glycemic index (HGI) were defined as GI<50 and GI≥50, respectively. The LGI diet group was instructed to select carbohydrate containing foods from a list of LGI grains, fruits, vegetables, and dairy, which was provided by the study researchers. Moreover, a list of prohibited HGI foods was given to LGI group members. We used a food exchange list to select foods in the meat and fat groups. We used several nutritional recommendations that emphasized limiting foods rich in fat, fast foods, French fries, fried foods, industrial beverages, and unhealthy fats. Participants were also recommended to drink 1.5-2 liters of water per day, consume a variety of fruits and vegetables and eat low-fat dairy and whole grains. The HNRD group's diet was based on these guidelines. All participants were asked to fill out a one-day food diary and a one-day physical activity record for 3 work days and one weekend day. The completed records were reviewed by trained staff. The participants were asked to complete food diaries based on household portion sizes. We converted quantities reported using household measures (e.g., a cup of milk) to grams [29]. Grams of foods were analyzed by Nutritionist IV to obtain the nutrient and energy content of the foods reported. Comparisons between macronutrient distribution as a percentage of energy in the prescribed diets and reported food records were used to check compliance in the HNRD group. The criterion for compliance in the LGI group was GI < 50, calculated from the food diaries. To extract GI values, we referred to the Iranian GI table [30]. The

To extract GI values, we referred to the Iranian GI table [30]. The international table of GI [31] was used for GI values not reported on the Iranian-specific table. The GI for foods not included in the Iranian or the international table was estimated using the GI for the most similar food. To calculate the mean GI for diet, we used the reported formula [32].

Biochemical measurements

One blood sample was taken after 12 h of fasting in the early morning. After coagulation, we centrifuged blood samples at $3\,000\,\times g$ for $10\,\text{min}$ to separate the serum. FBS was measured by an enzymatic colorimetric method based on glucose oxidase activity (Pars Azmoon, Tehran, Iran). Insulin was assessed by enzyme-linked immunosorbent assay (ELISA) (Monobind Inc, Costa Mesa, CA, USA). Standard sandwich ELISA technology was used for measuring adiponectin and IL-6 (Boster Biological Technology, USA). Intra- and interassay variations for IL-6 were less than 6 and 8%, respectively. The Immunoturbidimetry method was used to measure hs-CRP with a polyclonal antibody (Pars Azmoon Inc). For hs-CRP, the interassay and intra-assay variation was 5.1-10.0% and 4.3-6.1%, respectively. To estimate insu-

Table 1 Percent changes of fasting blood sugar, insulin, adiponectin, and inflammatory factors among adolescents in the low glycemic index (LGI) and healthy nutrition recommendations diet (HNRD) groups following a 10 week intervention.

Variables	Low glycemic index (LGI) group ¹ (n=25)	Healthy nutrition recommendations diet (HNRD) group ² (n = 25)	p ³	Model 1 ⁴	Model 2 ⁵
Fasting blood sugar	5.54 ± 2.45^6	4.24±2.20	0.69	0.81	0.97
Serum insulin	1.59 ± 1.95	1.67±2.03	0.96	0.37	0.32
HOMA	2.54±1.98	2.25±2.08	0.90	0.83	0.33
QUICKI	-0.00 ± 1.26	-0.75±1.96	0.74	0.57	0.43
Serum adiponectin	22.04±24.45	46.30±24.31	0.48	0.25	0.29
hs-CRP	-16.21±8.48	27.96±24.16	0.09	0.04	0.08
IL-6	-74.87±6.14	-70.27±6.92	0.62	0.84	0.48

HOMA: Homeostasis model assessment; QUICKI: Quantitative insulin sensitivity check index; hs-CRP: high sensitivity C-reactive protein; IL-6: Interleukin 6

lin resistance, homeostasis model assessment (HOMA) and the quantitative insulin sensitivity check index (QUICKI) were calculated with reported formulas [32].

Statistical analysis

The Kolmogorov-Smirnov test and histogram curves were used to test if the variables were normally distributed. Results showed that the distribution of adiponectin, IL-6 and percent changes of insulin, HOMA, hs-CRP, and IL-6 were not normal. Therefore, the geometric mean was used for these variables. Means of nutrient intake, baseline values, final values and percent changes were compared between the 2 groups with the Student's t-test. We used the following formula to calculate percent changes: [(E-B)/B] × 100 where baseline and final values were represented by B and E; respectively. Within group comparisons between baseline and final values were calculated with paired t-tests. We also used repeated measures ANOVA to calculate ptime, pgroup, $p_{time \times group}$ and $p_{time \times age}$. An intention-to-treat analysis was also performed. All variables are presented as mean ± SEM. All analyses were carried out using SPSS 20 (SPSS Inc) statistical software.

Results

V

The study procedure is presented in **Fig. 15**. Among 50 enrolled adolescents, 9 subjects withdrew due to poor compliance, change in phone number, or other reasons. Finally, 41 remained in the study. Using an intention-to-treat analysis, we included the data for all 50 subjects in the statistical analysis. As mentioned, the comparison between macronutrient distribution as a percentage of energy in the prescribed diets and reported food records were used as a compliance check in the HNRD group. No differences were observed between macronutrient composition between the prescribed and the consumed diets. A glycemic index of less than 50 was used as the compliance criterion in the LGI group. Analysis of food records demonstrated that the mean±SEM of GI in the LGI group was 42.67±0.67.

Baseline characteristics for adolescents in the LGI group and the HNRD group are displayed in **Table 1S**. Mean age in the LGI group was lower than in the HNRD group (p=0.03). There was no sig-

nificant difference in physical activity level between the 2 groups (p=0.43). Moreover, the percent of overweight subjects in the HNRD group was significantly higher than in the LGI group. Although weight decreased in both groups, changes were not different between the 2 groups. Analysis of food diaries showed that nutrient intake including energy intake, carbohydrate, protein, fat, fiber, zinc, vitamin E, vitamin B₃, folate, vitamin C, vitamin B₆, and vitamin D were not significantly different between the 2 groups (Table 1). Furthermore, dietary GI in the LGI group was lower than 42.67±0.067. According to physical activity records, there was not a significant difference in physical activity levels between the 2 groups (p = 0.43). We observed significant weight reduction in both groups (p<0.001 for both). The means of the insulin resistance indices, adiponectin and inflammatory factors, before and after the 10 weeks of the intervention are illustrated in • Table 2. Although average hs-CRP in the LGI group (p=0.002) and IL-6 in both groups (p<0.001 for both) decreased significantly after the 10 week intervention, other variables did not differ significantly within each group. The comparison between baseline and final values failed to show a significant difference between the 2 groups. Analyses did not demonstrate significant interactions except for a time \times age interaction for serum adiponectin concentration (p < 0.001). Percent changes in insulin resistance indices, serum adiponectin concentration, and inflammatory factors are presented in • Table 1. We compared percent changes in 2 different statistical models. In model 1 (adjusted for age), a significant reduction in hs-CRP was observed (p = 0.04). We did not observe any significant differences in other variables. In model 2 (adjusted for age, physical activity, weight change and the distribution of obese subjects in the 2 groups), a marginal decrease was observed in hs-CRP

Discussion

 $\overline{\mathbf{v}}$

As our results show, although changes in insulin resistance indices and adiponectin were not significantly different between the 2 groups, a marginally significant decrease in hs-CRP was observed in the LGI group compared to the HNRD group after adjusting for confounding variables. To our knowledge, this is

(p=0.08). Differences were not significant for the other variables.

¹GI < 50 was considered LGI

²Healthy nutritional recommendations included avoidance of foods rich in fat, fast foods, French fries, fried foods, industrial beverages and unhealthy fats as well as advice to drink 1.5–2 liters of water, consume more amount of diverse fruits and vegetables, and eat low-fat dairy and whole grains

³ p-values show percent changes between the 2 groups (independent samples t-tests were used for calculating p-values)

⁴Adjusted for age

⁵ Adjusted for age, physical activity, weight change, and the distribution of obese subjects in the 2 groups

⁶ Variables are presented as mean ± SEM except for insulin and HOMA, which are presented as geometric mean ± SEM

Table 2 Fasting blood sugar, insulin, serum adiponectin concentration, and inflammatory factors among adolescents in the low glycemic index (LGI) and healthy nutrition recommendations diet (HNRD) groups following a 10-week intervention.

Variables	Low glycemic index (LGI) group ¹ (n=25)	Healthy nutrition recommendation diet (HNRD) group ² (n=25)	p overall ³	P _{time} ⁴	P _{group} ⁵	P _{time×group254} 6	P _{time×age} ⁷		
Fasting blood sugar (mg/dl)									
Before	89.88 ± 1.91	91.86±1.67	0.44	0.58	0.24	0.94	0.70		
After	93.63 ± 0.98	94.99 ± 1.35	0.41						
p ⁸	0.07	0.11	-						
Serum insulin (µIU/mI)									
Before	20.30 ± 2.31	16.50 ± 1.48	0.17	0.30	0.26	0.22	0.26		
After	18.03 ± 1.43	15.93 ± 1.59	0.33						
p 8	0.10	0.66	-						
HOMA									
Before	4.55 ± 0.54	3.77 ± 0.34	0.23	0.28	0.36	0.29	0.25		
After	4.20±0.35	3.77 ± 0.40	0.42						
p 8	0.29	0.99							
QUICKI									
Before	0.31 ± 0.00	0.32 ± 0.01	0.31	0.50	0.31	0.48	0.53		
After	0.31 ± 0.00	0.32 ± 0.00	0.20						
p 8	0.46	0.41							
Serum adiponectin (ng/ml)									
Before	5.86 ± 1.15	3.98 ± 1.14	0.06	<0.001	0.04	0.03	<0.001		
After	3.95 ± 1.16	3.81 ± 1.16	0.86						
p ⁸	0.08	0.81	-						
hs-CRP (mg/l)									
Before	3.97 ± 0.35	3.48 ± 0.39	0.36	0.67	0.79	0.19	0.53		
After	2.62 ± 0.23	2.89±0.31	0.49						
p ⁸	0.002	0.25	-						
IL-6 (pg/ml)									
Before	74.44±1.33	60.86±1.26	0.59	0.09	0.33	0.37	0. 21		
After	10.21 ± 1.27	8.38 ± 1.28	0.57						
p ⁸	< 0.001	<0.001	-						

HOMA-IR: Homeostasis model assessment; QUICKI: Quantitative insulin sensitivity check index; hs-CRP: high sensitivity C-reactive protein; IL-6: Interleukin 6

the first study to examine the effects of an LGI diet on insulin resistance indices, serum adiponectin concentration and inflammatory markers simultaneously in overweight and obese female adolescents.

Although the effekt of GI on insulin resistance has been evaluated previously, reported results have been inconsistent. Several studies reported a beneficial effekt of LGI diets on insulin resistance among adolescents and children [19,22,23]. LGI foods are absorbed and digested slowly and therefore provide a prolonged full state in which free fatty acid release is suppressed. Therefore, improvements observed in glucose tolerance and insulin sensitivity may be observed following an LGI diet [33]. However, this beneficial effekt has also been observed following a hypocaloric HGI diet [24]. Based on those reported results, it is plausible that the effekt of calorie restriction is more important than dietary GI [24]. Because study subjects were adolescents, we prescribed moderate calorie restriction only among obese participants. Therefore, this might be the reason for not detecting a significant improvement in insulin resistance.

The evidence regarding the effects of GI on inflammatory factors among adolescents has been limited. In previous studies, a significant drop in inflammatory markers has been reported for both hypocaloric LGI and hypocaloric HGI diets in children [23,24]. In a longitudinal study conducted with 22 obese girls and boys, hs-CRP decreased after consumption of both hypocaloric LGI (GI=60) and hypocaloric HGI (GI=90) diets [24]. Although we reported a similar reduction in hs-CRP for the LGI group, there are several differences between the aforementioned study and our intervention, including participant gender, the definition of LGI and the degree of caloric restriction. Moreover, we observed a remarkable decrease in IL-6 in the LGI group as well as in hs-CRP in within-group analyses. On the other hand, our results show greater reduction in hs-CRP in the LGI group compared to the HNRD group, independent of age and weight change. This implies that weight change did not mediate the effekt of LGI diet on inflammation. HGI foods induce higher glucose concentration [30]. Glukose has been considered a stimulating factor for the expression of inflammatory markers [34].

¹GI < 50 was considered LGI

² Healthy nutritional recommendations included avoidance of foods rich in fat, fast foods, French fries, fried foods, industrial beverages, and unhealthy fats as well as advice to drink 1.5–2 liters of water, consume higher quantities of diverse fruits and vegetables, and eat low-fat dairy and whole grains

³ p-values show comparisons of baseline and final values between the 2 groups (independent sample t-tests were used for calculating p-values)

⁴p-values show the effekt of time (analysis of covariance was used for calculating p-values)

⁵ p-values show the effekt of grouping (analysis of covariance was used for calculating p-values)

 $^{^{6}}$ p-values show the time * group interaction (analysis of covariance was used for calculating p-values)

⁷p-values show the time * age interaction (analysis of covariance was used for calculating p-values)

⁸ p-values show comparisons of baseline and final values within each group (paired sample t-tests were used for calculating p-values)

⁹ Variables are presented as mean ± SEM except for adiponectin and IL-6, which are presented as geometric mean ± SEM

Further, the pattern of methylation at promoters of inflammatory genes is permanently changed by a temporary increment in glucose concentration [34].

Although several studies have examined the effects of GI on adiponectin in animals [35–37], limited research has been reported from clinical trials in humans. Mitsuhashi et al. [35] did not observe any difference in adiponectin concentration between LGI and HGI groups in dogs. However, this conclusion was inconsistent with studies by van Schothorst et al. and Pawlak et al. [36,37], which were conducted on mice. In a human study, the comparison between 2 meals with different GI showed that serum adiponectin concentration increased following consumption of an LGI meal [32]. Unlike that study, our intervention focused on the whole diet rather than a single meal. Further, participant age and anthropometric status in the present study were also distinct from that study.

Iranians do not consume HGI foods as staple foods because, as reported in the Iranian native GI table [30], Iranian staple foods (rice and white bread) are not HGI [38,39]. At the same time, for ethical reasons we could not prescribe an HGI diet in a sample of overweight and obese adolescents. Therefore, we could not compare the effects of an LGI diet on insulin resistance and inflammatory markers to an HGI diet in the present study. Another limitation of current study is that there was no biochemical indicator used to assess adherence to the diet. Also, we were unable to measure physical activity using a precise method such as accelerometry. Moreover, we did not match intervention groups for percent of obese subjects.

Statistically sufficient sample size, similarity in age and gender of participants and maintaining the same macro/micro nutrient intake between 2 groups are strengths of the current study.

In conclusion, changes in insulin resistance indices and serum adiponectin concentration were not significantly different between the 2 groups studied. Moreover, compliance with an LGI diet may have favorable effekt on inflammation among overweight and obese adolescent girls.

Acknowledgements

 $\overline{\mathbf{w}}$

This study was funded by a grant from the Food Security Research Center and Department of Community Nutrition, School of Nutrition and Food Science, Isfahan University of Medical Sciences.

Conflict of Interest

V

The authors declare no conflict of interest.

Affiliations

- ¹ Food Security Research Center, Isfahan University of Medical Sciences and Department of Community Nutrition, School of Nutrition and Food Sciences, Isfahan University of Medical Sciences, Isfahan, Iran
- ² Pediatrics Department, Child Growth and Development Research Center, Research Institute for Primordial Prevention of Non Communicable Disease, Isfahan University of Medical Sciences, Isfahan, Iran
- ³ Department of Pediatrics, Child Growth and Development Research Center, and School of Medicine, Isfahan University of Medical Sciences, Isfahan, Iran
- ⁴ Department of Pediatric Endocrinology and Metabolism Diseases, Endocrinology and Metabolism Research Center, School of Medicine, Isfahan University of Medical Sciences, Isfahan, Iran
- ⁵ Food Security Research Center, Isfahan University of Medical Sciences and Department of Community Nutrition, School of Nutrition and Food Sciences, Isfahan University of Medical Sciences, Isfahan, Iran

- ⁶ Department of International Health, Johns Hopkins Bloomberg, School of Public Health, Baltimore, USA
- ⁷ Department of Community Nutrition, School of Nutritional sciences and dietetics, Tehran University of Medical Sciences, Tehran, Iran
- ⁸ Department of Clinical Nutrition, School of Nutritional Sciences and Dietetics, Tehran University of Medical Sciences, Tehran, Iran

References

- 1 *Chou HH, Hsu LA, Liu CJ, Teng MS, Wu S, Ko YL.* Insulin resistance is associated with C-reactive protein independent of abdominal obesity in nondiabetic Taiwanese. Metabolism 2010; 59: 824–830
- 2 Mehrabian F, Khani B, Kelishadi R, Kermani N. The prevalence of metabolic syndrome and insulin resistance according to the phenotypic subgroups of polycystic ovary syndrome in a representative sample of Iranian females. J Res Med Sci 2011; 16: 763–769
- 3 McKenney RL, Short DK. Tipping the balance: the pathophysiology of obesity and type 2 diabetes mellitus. Surg Clin North Am 201 91: 1139–1148 vii
- 4 Yaturu S, Daberry RP, Rains J, Jain S. Resistin and adiponectin levels in subjects with coronary artery disease and type 2 diabetes. Cytokine 2006; 34: 219–223
- 5 Long J, Su YX, Deng HC. Lipoapoptosis pathways in pancreatic β -cells and the anti-apoptosis mechanisms of adiponectin. Horm Metab Res 2014; 46: 722–727
- 6 *Jian L, Su YX, Deng HC.* Adiponectin-induced inhibition of intrinsic and extrinsic apoptotic pathways protects pancreatic β-cells against apoptosis. Horm Metab Res 2013; 45: 561–566
- 7 Niehoff AG, van Haeften TW, Onland-Moret NC, Elbers CC, Wijmenga C, van der Schouw YT. C-reactive protein is independently associated with glucose but not with insulin resistance in healthy men. Diabetes Care 2007; 30: 1627–1629
- 8 Barbarroja N, Lopez-Pedrera C, Garrido-Sanchez L, Mayas MD, Oliva-Olivera W, Bernal-Lopez MR, El Bekay R, Tinahones FJ. Progression from high insulin resistance to type 2 diabetes does not entail additional visceral adipose tissue inflammation. PLoS One 2012; 7: e48155
- 9 Izadi V, Farabad E, Azadbakht L. Serum adiponectin level and different kinds of cancer: a review of recent evidence. ISRN Oncol 2012 982769
- 10 Keshavarz SA, Nourieh Z, Attar MJ, Azadbakht L. Effect of soymilk consumption on waist circumference and cardiovascular risks among overweight and obese female adults. Int J Prev Med 2012; 3: 798–805
- 11 Al-Isa AN, Thalib L, Akanji AO. Circulating markers of inflammation and endothelial dysfunction in Arab adolescent subjects: reference ranges and associations with age, gender, body mass and insulin sensitivity. Atherosclerosis 2010; 208: 543–549
- 12 Corcoran MP, Lamon-Fava S, Fielding RA. Skeletal muscle lipid deposition and insulin resistance: effect of dietary fatty acids and exercise. Am | ClinNutr 2007; 85: 662–677
- 13 Kasim-Karakas SE, Tsodikov A, Singh U, Jialal I. Responses of inflammatory markers to a low-fat, high-carbohydrate diet: effects of energy intake. Am J ClinNutr 2006; 83: 774–779
- 14 Salehi-Abargouei A, Izadi V, Azadbakht L. The effect of low calorie diet on adiponectin concentration: a systematic review and meta-analysis. Horm Metab Res 2015; 47: 549–555
- 15 Wolever TM, Yang M, Zeng XY, Atkinson F, Brand-Miller JC. Food glycemic index, as given in glycemic index tables, is a significant determinant of glycemic responses elicited by composite breakfast meals. Am J Clin Nutr 2006; 83: 1306–1312
- 16 Wolever TM, Gibbs AL, Mehling C, Chiasson JL, Connelly PW, Josse RG, Leiter LA, Maheux P, Rabasa-Lohret R, Rodger NW, Ryan EA. The Canadian Trial of Carbohydrates in Diabetes (CCD), a 1-y controlled trial of low-glycemic-index dietary carbohydrate in type 2 diabetes: no effect on glycated hemoglobin but reduction in C-reactive protein. Am J Clin Nutr 2008; 87: 114–125
- 17 McMillan-Price J, Petocz P, Atkinson F, O'Neill K, Samman S, Steinbeck K, Caterson I, Brand-Miller J. Comparison of 4 diets of varying glycemic load on weight loss and cardiovascular risk reduction in overweight and obese young adults: a randomized controlled trial. Arch Intern Med 2006; 166: 1466–1475
- 18 Bulló M, Casas R, Portillo MP, Basora J, Estruch R, García-Arellano A, Lasa A, Juanola-Falgarona M, Arós F, Salas-Salvadó J. Dietary glycemic index/load and peripheral adipokines and inflammatory markers in elderly subjects at high cardiovascular risk. Nutr Metab Cardiovasc Dis 2013; 23: 443–450
- 19 Armeno ML, Krochik AG, Mazza CS. Evaluation of two dietary treatments in obese hyperinsulinemic adolescents. J Pediatr Endocrinol Metab 2011; 24: 715–722

- 20 Weyman-Daum M, Fort P, Recker B, Lanes R, Lifshitz F. Glycemic response in children with insulin-dependent diabetes mellitus after high- or low-glycemic-index breakfast. Am J Clin Nutr 1987; 46: 798–803
- 21 Gilbertson HR, Brand-Miller JC, Thorburn AW, Evans S, Chondros P, Werther GA. The effect of flexible low glycemic index dietary advice versus measured carbohydrate exchange diets on glycemic control in children with type 1 diabetes. Diabetes Care 2001; 24: 1137–1143
- 22 Fajcsak Z, Gabor A, Kovacs V, Martos E. The effects of 6-week low glycemic load diet based on low glycemic index foods in overweight/obese children-pilot study. J Am CollNutr 2008; 27: 12-21
- 23 Iannuzzi A, Licenziati MR, Vacca M, De Marco D, Cinquegrana G, Laccetti M, Bresciani A, Covetti G, Iannuzzo G, Rubba P, Parillo M. Comparison of two diets of varying glycemic index on carotid subclinical atherosclerosis in obese children. Heart Vessels 2009: 24: 419–424
- 24 Parillo M, Licenziati MR, Vacca M, De Marco D, Iannuzzi A. Metabolic changes after a hypocaloric, low-glycemic-index diet in obese children. J Endocrinol Invest 2012; 35: 629–633
- 25 Azadbakht L, Esmaillzadeh A.. Dietary diversity score is related to obesity and abdominal adiposity among Iranian female youth. Public Health Nutr 2011; 14: 62–69
- 26 World Health Organization. BMI-for-age GIRLS [cited September 2011]. Available from:URL http://www.who.int/entity/growthref/bmifa_girls_5_19years_per.pdf
- 27 Fleiss JL. The design and analysis of clinical experiments. London: Wiley 1986; 263–271
- 28 Institute of medicine (US). Energy. In: Dietary reference intakes for energy, carbohydrate, fiber, fat, fatty acids, cholesterol, protein, and amino acids. 1st ed. Washington: The National Academies Press; 2005 217
- 29 Ghaffarpour M, Houshiar-Rad A, Kianfar H. The manual for household measures, cooking yield factors and edible portion of foods. Tehran: Keshaverzi Press; 1999 in Farsi
- 30 Azam-Taleban F, Esmaeili M. Glycemic index of Iranian foods. 1st ed.Tehran: National Nutrition and Food Technology Research Institute: 1999 in Farsi

- 31 Foster-Powell K, Holt SH, Brand-Miller JC. International table of glycemic index and glycemic load values: 2002. Am J ClinNutr. 2002; 76: 5–56
- 32 deRougemont A, Normand S, Nazare JA, Skilton MR, Sothier M, Vinoy S. Beneficial effects of a 5-week low-glycaemic index regimen on weight control and cardiovascular risk factors in overweight non-diabetic subjects. Br J Nutr 2007; 98: 1288–1298
- 33 Nilsson AC, Ostman EM, Holst JJ, Björck IM. Including indigestible carbohydrates in the evening meal of healthy subjects improves glucose tolerance, lowers inflammatory markers, and increases satiety after a subsequent standardized breakfast. J Nutr 2008; 138: 732–739
- 34 Gögebakan O, Kohl A, Osterhoff MA, van Baak MA, Jebb SA, Papadaki A, Martinez JA, Handjieva-Darlenska T, Hlavaty P, Weickert MO, Holst C, Saris WHM, Astrup A, Pfeiffer AFH. MD on behalf of Genes. Effects of weight loss and long-term weight maintenance with diets varying in protein and glycemic index on cardiovascular risk factors: the diet, obesity, and genes. (DiOGenes) study: a randomized, controlled trial. Circulation 2011; 124: 2829–2838
- 35 Mitsuhashi Y, Nagaoka D, Ishioka K, Bigley KE, Okawa M, Otsuji K, Bauer JE. Postprandial lipid-related metabolites are altered in dogs fed dietary diacylglycerol and low glycemic index starch during weight loss. | Nutr 2010; 140: 1815–1823
- 36 vanSchothorst EM, Bunschoten A, Schrauwen P, Mensink RP, Keijer J. Effects of a high-fat, low- versus high-glycemic index diet: retardation of insulin resistance involves adipose tissue modulation. FASEB J 2009; 23: 1092–1101
- 37 Pawlak DB, Kushner JA, Ludwig DS. Effects of dietary glycaemic index on adiposity, glucose homoeostasis, and plasma lipids in animals. Lancet 2004; 364: 778–785
- 38 Esmaillzadeh A, Azadbakht L. Food intake patterns may explain the high prevalence of cardiovascular risk factors among Iranian women. J Nutr 2008; 138: 1469–1475
- 39 Azadbakht L, Esmaillzadeh A. Dietary patterns and attention deficit hyperactivity disorder among Iranian children. Nutrition 2012; 28: 242–249